



Identification of the Top 10 Core Genes as Diagnostic and Therapeutic Targets in Hepatocellular Carcinoma (HCC)

Qiangsheng Wu¹, Yumei Yin², Hui Gao¹, Baoding Chen³, Jianjun Yu², Jian Zhang⁴, Donglai Zhu⁵, Fang Liu⁶, Guohong Ge⁷ and Ya-Mei Dang^{8*}

¹Department of Assay Development, Eoto-Biotech Co., Ltd, Shenzhen, Guangdong 518000, China.

²Department of Medical Ultrasound, The Third People's Hospital of Zhenjiang Affiliated Jiangsu University, Zhenjiang, Jiangsu 212005, China.

³Department of Medical Ultrasound, The Affiliated Hospital of Jiangsu University, Zhenjiang, Jiangsu 212001, China.

⁴Department of Radiology, The Affiliated Hospital of Jiangsu University, Zhenjiang, Jiangsu 212001, China.

⁵Department of Clinical Laboratory, The Third People's Hospital of Zhenjiang Affiliated Jiangsu University, Zhenjiang, Jiangsu 212005, China.

⁶Department of Computer, Anhui Medical University, Hefei, Anhui 230032, China.

⁷Department of Hepatosis, The Third People's Hospital of Zhenjiang Affiliated Jiangsu University, Zhenjiang, Jiangsu 212005, China.

⁸Department of Pathology, Gansu Provincial Hospital, Lanzhou, Gansu 730000, China.

Qiangsheng Wu, Yumei Yin and Hui Gao contributed equally to this work.

ABSTRACT

Lacking specific diagnostic markers for hepatocellular carcinoma is the critical reason for its increasing mortality. In this study, a microarray containing 24 hepatocellular carcinoma patients' tumor tissues were compared to 8 normal liver tissues to screen out the significant abnormal expressed genes. The potential interactions and mechanisms of these genes in promoting hepatocellular carcinoma were analyzed through bioinformatics analysis. The results showed that the functions of these 1495 differently expressed genes enriched in cell cycle, oxidation-reduction and drug metabolic process. Additionally, the top 10 core genes of these differently expressed genes were mainly participated in cell cycle and strongly correlated with worse prognosis of survival, which may be potential diagnostic and therapeutic targets of HCC.

INTRODUCTION

Hepatocellular carcinoma (HCC) is a lethal malignancy of the liver. Primary liver cancer (85% was HCC) was the third leading cause of mortal cancer in the worldwide. Notably, 49.3% of the new cases and 58.3% of the deaths

were occurred in Asia (Sung *et al.*, 2021). HCC is associated with potentially alterable risk factors, such as excess body weight, alcohol abuse, hepatitis B virus (HBV), hepatitis C virus, nonalcohol fatty liver disease (Islami *et al.*, 2018). As a high malignancy with insidious onset, invasive fast growing, high recurrence rate and fatality, HCC is a major public health problem especially in developing countries (McGlynn *et al.*, 2021).

Effective and sensitive diagnostic and curative interventions available for HCC can serve to reduce the future burden and suffering of patients (Yang *et al.*, 2019). Currently, diagnosis available for HCC screening relies on radio graphic tests and serological markers. However, most small nodules less than 2cm are extremely insensitive to be imaging detected, as well as the sensitivity of the serum golden marker AFP only ranges of 21%-64%

* Corresponding author: yameidanggs@126.com
0030-9923/2022/0001-0001 \$ 9.00/0



Copyright 2022 by the authors. Licensee Zoological Society of Pakistan.

This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Article Information

Received 15 April 2022

Revised 08 May 2022

Accepted 26 May 2022

Available online 30 September 2022
(early access)

Authors' Contribution

QSW, YMY and HG performed data analysis work and drafted in writing the manuscript. BDC and JJY assisted in collecting clinical and research advances. DLZ and JZ revised the manuscript. FL assisted in computer data processing. GHG and YMD designed the study and assisted in writing the manuscript. All authors agreed to be accountable for all aspects of the work.

Key words

Hepatocellular carcinoma, Diagnostics, Therapeutic targets, Bioinformatic analyses, Cell cycle

(even lower with small lesions) (Yang and Heimbach, 2020). The unfortunate truth is that it is often too late stages when patients have become symptomatic and been diagnosed, while no effective treatment would improve survival virtually (Singal *et al.*, 2020; Yang *et al.*, 2019). There is absolutely a need for higher sensitive and specific diagnostic markers for HCC.

Recently, bioinformatics analysis based on microarrays and high throughput sequencing technologies has been widely used to identifying differentially expressed genes (DEGs) and pathways related to the occurrence and developments of various cancers (Zhang *et al.*, 2021). In this article, we provide comprehensive biological analysis of new potential targets of HCC and their functions in promoting and regulating tumor progression. Which hopefully will have unique contributions to ensure optimal treatments and outcomes for patients with HCC.

MATERIALS AND METHODS

Microarray and differentially expressed genes analysis

The original data of GSE101685 microarray was downloaded from GEO datasets (<https://www.ncbi.nlm.nih.gov/geo/>). The gene expression profiles of 8 normal liver and 24 hepatocellular carcinoma tissues were selected and compared. Genes with adjusted $|\log_2FC| \geq 1$ and p value < 0.05 were identified as differentially expressed genes (DEGs).

Functional enrichment analysis

The differentially expressed gene list was uploaded to databases of DAVID 6.8 (<https://david.ncifcrf.gov/home.jsp>) and Metascape (<https://metascape.org>) separately (Zhou *et al.*, 2019a). These two databases provided a comprehensive set of functional annotation for the biological meaning, which contained the Gene Ontology (GO) enrichment analysis and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analysis. In addition, the GO enrichment analysis included biological processes (BP), cellular components (CC), and molecular function (MF).

Neighbor genes and interaction network analysis

The differentially expressed gene list was uploaded to an open source software platform (Cytoscape) (Wardle-Farley *et al.*, 2010). Plugin STRING was used for analyzing the interaction network of DEGs. Plugin mCODE was used for finding the most highly interconnected region in DEGs. Plugin cytoHubba was used for exploring the most important nodes/hubs.

Top 10 core genes analysis (Shannon *et al.*, 2003)

Interaction network of neighbor genes associated

with the top 10 core genes was revealed on GeneMANIA (<http://genemania.org/>). Genetic alteration analysis of the top 10 core genes in HCC was explored from cBioPortal (www.cbioportal.org) based on TCGA database.

Aberrant expression analysis

Transcriptional levels of the top 10 core genes in 371 HCC and 50 normal tissues were explored and compared using UALCAN. Student's t test was used to generate a p value. ****, $p < 0.0001$.

Prognostic value analysis (Tang *et al.*, 2017)

Correlative prognostic analysis of the top 10 core genes expression was performed using a Kaplan-Meier curve on GEPIA (<http://gepia.cancer-pku.cn/index.html>). Disease-free survival and overall survival curves were analyzed separately.

RESULTS

Gene expression profile of normal and tumor tissues of HCC

The microarray GSE101685 containing 8 normal liver tissues and 24 HCC patients' tumor tissues was selected and showed the gene expression profile (Fig. 1A). 1495 different expressed genes were obtained with both the adjusted $|\log_2FC| \geq 1$ and $p < 0.05$. Compared with the normal tissue, these genes different-expressed in HCC tissue include 650 up-regulated genes and 845 down-regulated genes (Fig. 1B).

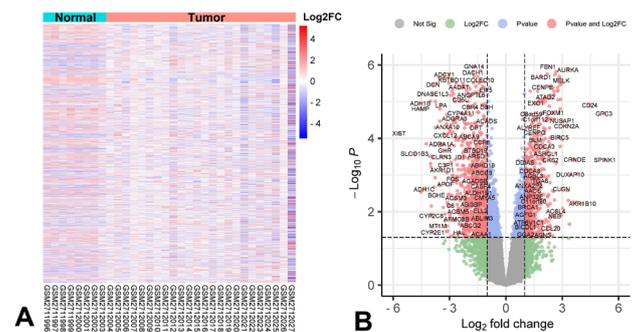


Fig. 1. Different-expressed genes in HCC tumor tissue. (A) The gene expression profile in normal liver tissue and HCC tissue. (B) Statistically significant up-regulated or down-regulated expressed genes (red) in HCC tissue.

Functional enrichment analysis of DEGs in HCC

DAVID 6.8 and Metascape were utilized to investigate the functions of these differentially expressed genes. Figure 2A, B showed the most highly enriched GO and KEGG items of the significant up-regulated expressed

genes. Among the most highly enriched functions in the BP, CC and MF category or in KEGG pathways. Cell cycle was the most central functional item of all the up-regulated expressed genes. Figure 2C, D showed the most highly enriched GO and KEGG items of the significant down-regulated expressed genes. All these genes were mainly involved in oxidation-reduction and drug metabolic process.

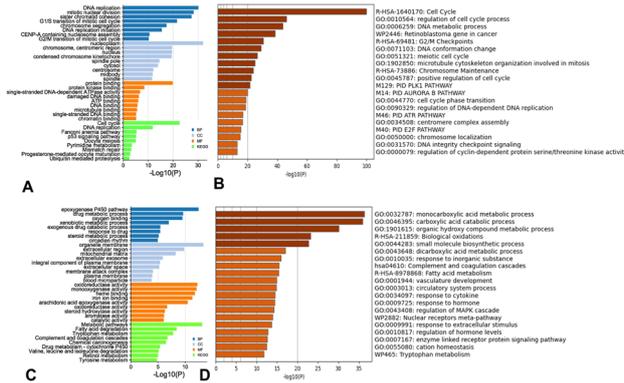


Fig. 2. Enrichment analysis of the DEG in HCC. Bar plot of GO (BP, CC, MF) and KEGG enriched items of up-regulated genes analyzed by DAVID 6.8 (A) and Metascape (B). Bar plot of GO (BP, CC, MF) and KEGG enriched items of down-regulated genes analyzed by DAVID 6.8 (C) and Metascape (D).

The interaction network, mCODE and the top 10 core genes of the DEGs

To better understand the correction between these DEGs and HCC, the PPI network and the most significant mCODE component were analyzed (Fig. 3A, B). Then the Cytoscape plugin cytoHubba further ranked and showed the top 10 core genes. These core genes included *NCAPG*, *NUF2*, *CCNB1*, *KIF11*, *BUB1B*, *RRM2*, *TTK*, *CCNB2*, *CCNA2* and *MELK* (Fig. 3C).

Neighbor gene network, functional enrichment and genetic alteration of the top 10 core genes

We further performed a comprehensive analysis of these 10 core genes. Firstly, Gene MANIA showed the neighbor gene network and revealed the functions of these core genes were primarily related to mitotic nuclear division, chromosome segregation and cell cycle checkpoint (Fig. 4A). While the functional enrichment was checked by Cytoscape. As expected, the core genes focused on cell cycle (Fig. 4B). Subsequently, provisional datasets of TCGA were utilized to analyze the genetic alterations. *NCAPG*, *NUF2*, *CCNB1*, *KIF11*, *BUB1B*, *RRM2*, *TTK*, *CCNB2*, *CCNA2* and *MELK* were altered in

8%, 27%, 9%, 6%, 7%, 9%, 8%, 9%, 4% and 8% of the queried HCC samples. And mRNA high expression was the most common change in these samples (Fig. 4C).

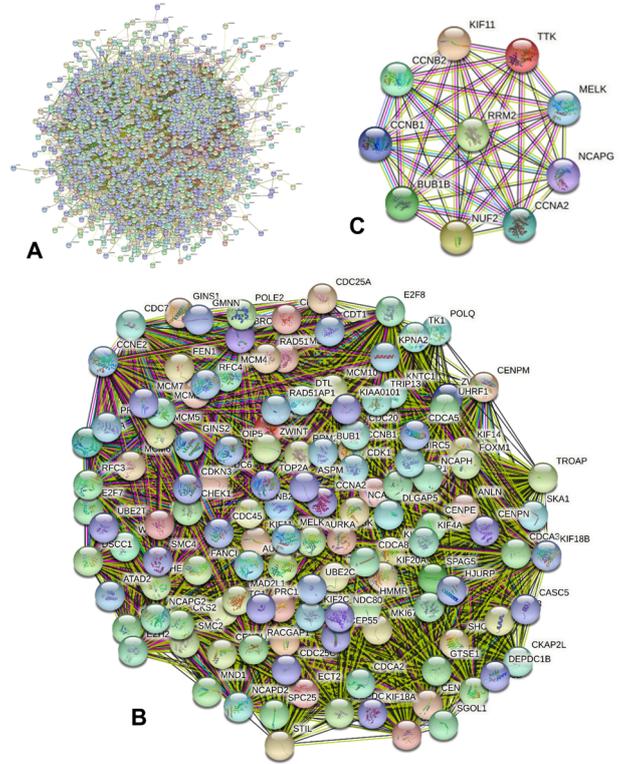


Fig. 3. Interaction network, mCODE and the top 10 core genes analyses of the DEGs in HCC. (A) Protein-protein interaction network of the DEGs in HCC (STRING). (B, C) The most significant mCODE and top 10 core genes of the DEGs in HCC (cytoscape).

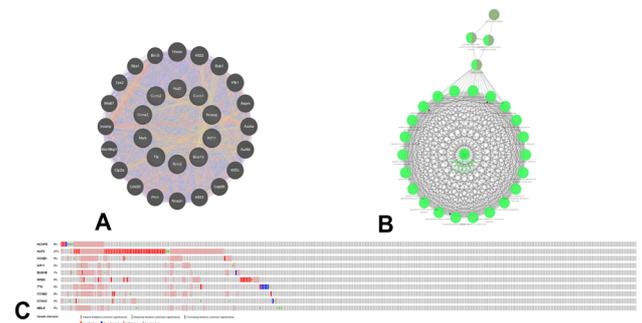


Fig. 4. Neighbor gene network, functional enrichment and genetic alteration of the top 10 core genes in HCC. Gene-gene interaction network (A) and the functional enrichment (B) of the top 10 core genes and their neighbor genes. (C) Summary of genetic alteration of the top 10 core genes in HCC.

Aberrant expression of the top 10 core genes in patients with HCC

To assess the expression levels of these top 10 core genes in larger sample size of HCC, the transcriptional levels of these genes in 371 HCC and 50 normal tissues were explored with UALCAN based on TCGA database (Fig. 5). As expected, the transcriptional levels of *NCAPG*, *NUF2*, *CCNB1*, *KIF11*, *BUB1B*, *RRM2*, *TTK*, *CCNB2*, *CCNA2* and *MELK* in HCC tissues were significantly elevated (****, $p < 0.0001$). Which indicated these genes have potential applications in the field of HCC diagnosis.

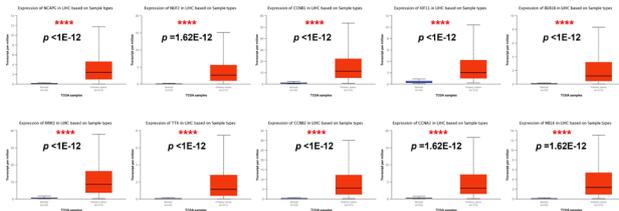


Fig. 5. The transcriptional levels of the top 10 core genes in HCC. Compared the transcription of *NCAPG*, *NUF2*, *CCNB1*, *KIF11*, *BUB1B*, *RRM2*, *TTK*, *CCNB2*, *CCNA2* and *MELK* of HCC tissues with the normal tissues (UALCAN). LIHC represents liver hepatocellular carcinoma. ****, $p < 0.0001$.

The prognostic value of the top 10 core genes in patients with HCC

To evaluate the value of differentially expressed core genes in the progression of HCC, we assessed the correlation between the top 10 core genes and clinical outcome using GEPIA. Disease-free survival and overall survival curves are presented in Figure 6. HCC patients with low transcriptional levels of *NCAPG* ($p=0.0024$), *NUF2* ($p=0.0038$), *CCNB1* ($p=0.0000028$), *KIF11* ($p=0.00024$), *BUB1B* ($p=0.00072$), *RRM2* ($p=0.00032$), *TTK* ($p=0.0088$), *CCNB2* ($p=0.0064$), *CCNA2* ($p=0.0037$) and *MELK* ($p=0.0014$) were significantly associated with longer disease-free survival (Fig. 6A). HCC patients with low transcriptional levels of *NCAPG* ($p=0.00097$), *NUF2* ($p=0.0002$), *CCNB1* ($p=0.00015$), *KIF11* ($p=0.00061$), *BUB1B* ($p=0.0028$), *RRM2* ($p=0.00058$), *TTK* ($p=0.0015$), *CCNA2* ($p=0.0037$) and *MELK* ($p=0.0015$) were significantly associated with better overall survival, except the *CCNB2* (Fig. 6B). All these data demonstrate the differently expressed top 10 core genes may play significant roles and be potential therapeutic targets in HCC.

DISCUSSION

Although amount of basic and clinical studies has been performed to reveal the underlying molecular mechanism

of HCC, the HCC-related genes have yet to be identified to elucidate the correlated with the cancer susceptibility, progression, and prognosis. This is mainly because the heterogeneity of the sample in independent studies or the single cohort nature in most of the previous studies. In this study, a microarray data containing 8 normal liver tissues and 24 HCC patients' tumor tissues was analyzed, and a total of 1495 DEGs (650 upregulated and 845 down regulated) were identified in HCC tissues compared with normal liver tissues. For the further investigation of these DEGs, GO function and KEGG pathway analysis of these DEGs were performed.

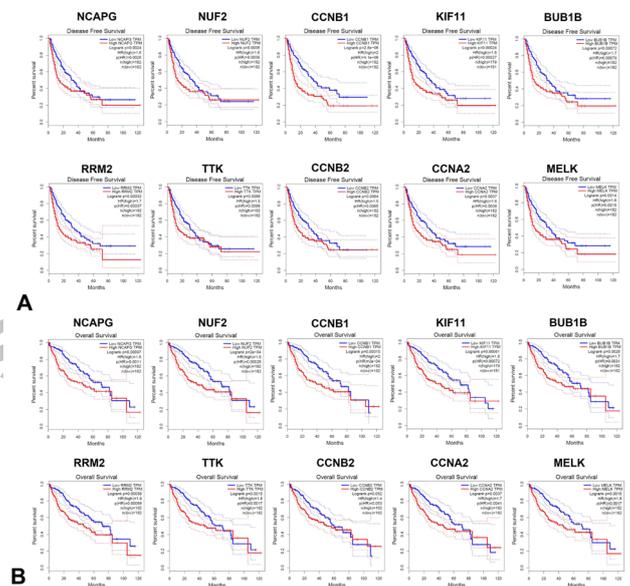


Fig. 6. The prognostic value of the top 10 core genes in patients with HCC. The disease-free survival (A) and overall survival (B) curves of *NCAPG*, *NUF2*, *CCNB1*, *KIF11*, *BUB1B*, *RRM2*, *TTK*, *CCNB2*, *CCNA2* and *MELK* in HCC.

GO analysis showed that upregulated DEGs were mainly enriched in cellular component groups, including nucleoplasm, chromosome, nucleus, and down-regulated DEGs were involved in organelle membrane, extracellular region, and mitochondrial matrix. Moreover, the KEGG pathways of upregulated DEGs were involved in cell cycle, DNA replication, Fanconi anemia pathway, and p53 signaling pathway, while the downregulated DEGs included metabolic pathways, fatty acid degradation, tryptophan metabolism, and complement and coagulation cascades. Afterward, the DEG PPI network complex was constructed, ten DEGs with the highest degrees of interaction in the PPI network complex were filtered.

To better understand the correction between these

DEGs and HCC, the neighbor Gene Network, functional enrichment, and genetic alteration of the top ten core genes was analyzed. The results revealed that the functions of these core genes were primarily related to mitotic nuclear division, chromosome segregation and cell cycle checkpoint. Finally, the aberrant expression of the 10 core genes shown that the transcriptional levels of *NCAPG*, *NUF2*, *CCNB1*, *KIF11*, *BUB1B*, *RRM2*, *TTK*, *CCNB2*, *CCNA2* and *MELK* were significantly elevated in HCC tissues. In addition, the patients with dysregulation of the ten core genes had a worse prognosis, except the *CCNB2*. Consulting the latest research on the top 10 core genes, we found these genes were significantly connected with multiple cancers.

NCAPG (non-SMC condensin I complex subunit G), a subunit of the condensin complex, is associated with transcription in mitotic chromosome condensation. The increasing evidence reported that aberrant expression of NCAPG in various tumors, the inhibition of NCAPG can reduce Wnt/ β -catenin signaling to impair the growth of endometrial cancer cells (Liu *et al.*, 2021). NCAPG-related stimulation could activate PI3K/Akt pathway in the development of cardia adenocarcinoma (Zhang *et al.*, 2020a). NCAPG has been found to be overexpressed in HCC compared with para-carcinoma tissue and is closely associated with poor prognosis and tumor immune infiltration in HCC patients with vascular (Liu *et al.*, 2017b). Recently study have reported that NCAPG was significantly associated with tumor cell proliferation and migration in HCC (Zhang *et al.*, 2018). Silencing NCAPG inhibits proliferation and induces apoptosis in HCC cells (Liu *et al.*, 2018). Nevertheless, the mechanism by which NCAPG promotes proliferation in HCC remains unknown. Interestingly, the overexpression of NCAPG is closely related to the overexpression of CCNB1, which is involved in the ten core genes of HCC (Wang *et al.*, 2019a).

NUF2, a component of NDC80 kinetochore complex is essential for kinetochore-microtubule attachment and chromosome segregation. The recent study demonstrated that high NUF2 transcript expression can be a good prognostic biomarker to predict early tumor recurrence post-surgical resection in HCC. And the inhibition of NUF2 reduced the proliferation, migration, and invasion of HCC cells in vitro (Wang *et al.*, 2019b). Nuf2 regulated cell apoptosis and proliferation by regulating the binding of centromere and spindle microtubules to achieve the correct separation of chromosomes (Hu *et al.*, 2015). The inhibition of NUF2 has been reported to reduce tumor cell growth in cancers of the brain, colon, liver, and pancreas (Huang *et al.*, 2014). However, the role of Nuf2 in the development of HCC remains uncertain.

CCNB1/CCNB2/CCNA2 as members of cyclin

family, act as recognized oncogenes in cervical cancer, breast cancer, colorectal cancer and pancreatic cancer (Sugimasa *et al.*, 2015). CCNB1 (cyclin B1) and CCNB2 (cyclin B2) can form complexes with cyclin-dependent kinase 1 (CDK1) to regulate the G2/M phases of the mammalian cell cycle, which plays an important role in the initiation of mitosis (Malumbres and Barbacid, 2005). As shown in recent studies, the mRNA expression levels of CCNB1 and CCNB2 were significantly higher in several types of cancer and were associated with poor prognosis (Ding *et al.*, 2014). Interestingly, in our study HCC patients with low transcriptional levels of CCNB2 associated with longer disease-free survival but not associated with overall survival. Knockdown of CCNB1 or CCNB2 could inhibit cell proliferation, invasion, migration, and G2/M phase progression of HCC cell lines and induced their apoptosis (Fang *et al.*, 2014). As a cyclin controlling the G1/S and G2/M phases in the cell cycle, CCNA2 has been reported to participate in HCC development (Yam *et al.*, 2002). The recent study indicated that overexpressed level of CCNA2 with other cell cycle regulators correlated with poor survival condition HCC subclass (Bai *et al.*, 2020), which was in agreement with our prognostic analysis results. Other studies shown that CCNA2 regulated cell cycle and promote antiapoptotic activity in HCC patients (Qian *et al.*, 2015).

Kinesin family member 11 (KIF11, also known as EG5), belongs to the kinesin-like protein family, is mainly involved in various kinds of spindle dynamics KIF11 functions as an oncogene in breast cancer, glioblastoma and oral cancer (Yang *et al.*, 2009). The recent research demonstrated that high KIF11 expression in HCC tissues, which is strongly associated with liver cirrhosis and tumor stages and can be considered as a biomarker for poor prognosis of HCC (Liu *et al.*, 2017a). Additionally, KIF11 interact with ASPM could promote the malignant progression of HCC via the Wnt/ β -catenin signaling pathway (Wu *et al.*, 2021). PRC1 plays a carcinogenic role by regulating KIF11 and promoting the occurrence and development of HCC (Chen *et al.*, 2016). The *in vivo* study demonstrated that overexpression of KIF11 causes genomic instability in tumor cells and prompts abnormal proliferation in mice (Castillo *et al.*, 2007).

BUB1B (BUB1 mitotic checkpoint serine/threonine kinase B) encodes a kinase involved in spindle checkpoint function. As a significant checkpoint of cell mitosis, BUB1B is involved in process of cell mitosis (Shin *et al.*, 2003), and upregulation of BUB1B is observed in various human malignancies (Ando *et al.*, 2010). Growing evidence shows that aberrant expression of BUB1B was highly involved in the tumorigenesis and development various cancers, for instance, overexpression of BUB1B

is related to gastric cancer (Hanks *et al.*, 2004), colon cancer (Abal *et al.*, 2007), brain tumor (Ding *et al.*, 2013), glioblastoma (Lee *et al.*, 2017) and breast cancer (Scintu *et al.*, 2007). Meanwhile, low expression of BUB1B associated metastasis and poorer survival have been found in colon adenocarcinoma (Shichiri *et al.*, 2002) and lung tumor (Park *et al.*, 2007). Recent study demonstrated that activation of FoxM1/BUB1B signaling pathway was vital for rhabdomyosarcoma growth and survival (Wan *et al.*, 2012). Nevertheless, we still know quite little about the functional roles and potential mechanisms of BUB1B in HCC.

Human ribonucleotide reductase (RR) is a heterotetrameric complex by two large subunits RRM1 (ribonucleotide reductase catalytic subunit M1) and two small subunits RRM2 (ribonucleotide reductase regulatory subunit M2) (Heidel *et al.*, 2007). RR maintains the homeostasis of nucleotide pools by converting ribonucleoside diphosphate to 20 deoxyribonucleoside diphosphate (Nordlund and Reichard, 2006). An-cancer expression profiling studies demonstrated that the expression of RRM2 is upregulated in multi types of cancers (Ding *et al.*, 2019; Shao *et al.*, 2013), dramatically enhances cellular invasiveness, angiogenesis, and metastasis (Zhou *et al.*, 1998). In HCC tissues the expression of RRM2 is upregulated compared with adjacent normal liver tissues (Wang *et al.*, 2010). The recent study revealed that the upregulation of RRM2 is closely associated with the poor prognosis of HCC patients, and RRM2 is enriched in the p53 signaling pathway (Zhou *et al.*, 2019b). Additionally, Several RRM2 inhibitors are currently clinically used for anticancer and antiviral therapy (Shao *et al.*, 2013). RRM2 siRNA transfection efficiently inhibited the cell proliferation of HCC cells (Yang *et al.*, 2020). The *in vivo* study shows that the RRM2 siRNA alone or in combination with adriamycin could inhibit xenografted or orthotopic tumor growth established in nude mice (Gao *et al.*, 2013).

TTK encodes a dual specificity protein kinase associated with cell proliferation. Differently expressed TTK may result in aberrant mitotic spindles and multiple transcript variants (Mills *et al.*, 1992). Accumulated evidence shown that overexpression of TTK could be detected in different types of cancer, including HCC (Jemal *et al.*, 2011; Xie *et al.*, 2017), glioblastoma (Xie *et al.*, 2017), esophageal cancer (Mizukami *et al.*, 2008), and breast cancer (Fan *et al.*, 2006). In our study, TTK was overexpressed in HCC tissues on mRNA levels and HCC patients with high expression levels of TTK had lower survival rates. The recent research had speculated that TTK in HCC carcinogenesis by promoting cell survival and invasion via activation of Akt/mTOR and MDM2/p53 signaling pathways, and the regulation of miR-21

via TGF- β (Liu *et al.*, 2015). In addition, TTK plays an essential role in proliferation and sorafenib resistance of HCC cells and could act as a biomarker and potential target for HCC treatment (Liang *et al.*, 2014).

Maternal embryonic leucine zipper kinase (MELK) is a unique member of the Snf1/AMPK kinase family. It belongs to a conservative cycle-dependent kinase (Li *et al.*, 2022). MELK plays an important role in cell cycle regulation, cell proliferation and other processes (Nakano *et al.*, 2005). MELK is highly expressed in a variety of human tumors, the recent study shown that E2F1/MELK/mTORC1/2 axis involved in the progression of endometrial carcinoma (EC) (Xu *et al.*, 2020). The inhibition of MELK could suppress the growth of Glioblastoma multiforme (GBM) via blockage AKT signals (Zhang *et al.*, 2020b). Numerous studies regarding the development of HCC provide evidence that variation in expression level of MELK is concerned with the development of HCC (Xia *et al.*, 2016). In our study, the analysis of HCC patients identified that mRNA level of MELK was highly overexpressed, which was correlated with early recurrence and poor overall survival. Further study demonstrated that silencing MELK inhibited the cell growth, invasion, stemness and tumorigenicity of HCC cells by inducing apoptosis and mitosis (Xia *et al.*, 2016).

Overall, by using a stepwise analysis of a microarray of 24 HCC tumor tissues and 8 normal liver tissues, we identified 1495 DEGs candidate genes (650 upregulated and 845 downregulated). Ten core genes with the highest interaction degrees were obtained, and these genes were significantly enriched in several pathways, mainly related to mitotic nuclear division, chromosome segregation and cell cycle checkpoint. The aberrant expression and survival analysis also showed a close interaction between these core genes and the poor prognosis of HCC.

In this study, the top 10 core genes *NCAPG*, *NUF2*, *CCNB1*, *KIF11*, *BUB1B*, *RRM2*, *TTK*, *CCNB2*, *CCNA2* and *MELK* might be important oncogenes of cell cycle in HCC. These findings could significantly enrich our understanding of the development and recurrence of HCC. Moreover, these genes could be therapeutic targets for HCC treatment. Future studies will be needed to further understand the underlying mechanisms of the concrete interaction of these genes.

ACKNOWLEDGMENTS

This work was supported by grant from Science and Technological Development Foundation of Zhenjiang (SH2019086, SH2020035, FZ2020050).

Statement of conflict of interest

The authors have declared no conflict of interest.

REFERENCES

- Abal, M., Obrador-Hevia, A., Janssen, K., Casadome, L., Menendez, M., Carpentier, S., Barillot, E., Wagner, M., Ansoorge, W., Moeslein, G., Fsihi, H., Bezrookove, V., Reventos, J., Louvard, D., Capella, G., and Robine, S.J.G., 2007. APC inactivation associates with abnormal mitosis completion and concomitant BUB1B/MAD2L1 up-regulation. *Gastroenterology*, **132**: 2448-2458. <https://doi.org/10.1053/j.gastro.2007.03.027>
- Ando, K., Kakeji, Y., Kitao, H., Iimori, M., Zhao, Y., Yoshida, R., Oki, E., Yoshinaga, K., Matumoto, T., Morita, M., Sakaguchi, Y., and Maehara, Y.J.C.s., 2010. High expression of BUBR1 is one of the factors for inducing DNA aneuploidy and progression in gastric cancer. *Cancer Sci*, **101**: 639-645. <https://doi.org/10.1111/j.1349-7006.2009.01457.x>
- Bai, X., Wang, W., Zhao, P., Wen, J., Guo, X., Shen, T., Shen, J., and Yang, X., 2020. LncRNA CRNDE acts as an oncogene in cervical cancer through sponging miR-183 to regulate CCNB1 expression. *Carcinogenesis*, **41**: 111-121. <https://doi.org/10.1093/carcin/bgz166>
- Castillo, A., Morse, H., Godfrey, V., Naeem, R., and Justice, M.J.C.R., 2007. Overexpression of Eg5 causes genomic instability and tumor formation in mice. *Cancer Res.*, **67**: 10138-10147. <https://doi.org/10.1158/0008-5472.CAN-07-0326>
- Chen, J., Rajasekaran, M., Xia, H., Zhang, X., Kong, S., Sekar, K., Seshachalam, V., Deivasigamani, A., Goh, B., Ooi, L., Hong, W., and Hui, K.J.G., 2016. The microtubule-associated protein PRC1 promotes early recurrence of hepatocellular carcinoma in association with the Wnt/ β -catenin signalling pathway. *Gut*, **65**: 1522-1534. <https://doi.org/10.1136/gutjnl-2015-310625>
- Ding, K., Li, W., Zou, Z., Zou, X., and Wang, C., 2014. CCNB1 is a prognostic biomarker for ER+ breast cancer. *Med. Hypoth.*, **83**: 359-364. <https://doi.org/10.1016/j.mehy.2014.06.013>
- Ding, Y., Hubert, C., Herman, J., Corrin, P., Toledo, C., Skutt-Kakaria, K., Vazquez, J., Basom, R., Zhang, B., Risler, J., Pollard, S., Nam, D., Delrow, J., Zhu, J., Lee, J., DeLuca, J., Olson, J., and Paddison, P.J.C.D., 2013. Cancer-Specific requirement for BUB1B/BUBR1 in human brain tumor isolates and genetically transformed cells. *Cancer Discov.*, **3**: 198-211. <https://doi.org/10.1158/2159-8290.CD-12-0353>
- Ding, Y., Zhong, T., Wang, M., Xiang, X., Ren, G., Jia, Z., Lin, Q., Liu, Q., Dong, J., Li, L., Li, X., Jiang, H., Zhu, L., Li, H., Shen, D., Teng, L., Li, C., and Shao, J., 2019. Integrative analysis reveals across-cancer expression patterns and clinical relevance of ribonucleotide reductase in human cancers. *Front. Oncol.*, **9**: 956. <https://doi.org/10.3389/fonc.2019.00956>
- Fan, C., Oh, D., Wessels, L., Weigelt, B., Nuyten, D., Nobel, A., van't Veer, L., and Perou, C., 2006. Concordance among gene-expression-based predictors for breast cancer. *N. Engl. J. Med.*, **355**: 560-569. <https://doi.org/10.1056/NEJMoa052933>
- Fang, Y., Yu, H., Liang, X., Xu, J., and Cai, X., 2014. Chk1-induced CCNB1 overexpression promotes cell proliferation and tumor growth in human colorectal cancer. *Cancer Biol. Ther.*, **15**: 1268-1279. <https://doi.org/10.4161/cbt.29691>
- Gao, J., Chen, H., Yu, Y., Song, J., Song, H., Su, X., Li, W., Tong, X., Qian, W., Wang, H., Dai, J., and Guo, Y.J.B., 2013. Inhibition of hepatocellular carcinoma growth using immunoliposomes for co-delivery of adriamycin and ribonucleotide reductase M2 siRNA. *Biomaterials*, **34**, 10084-10098. <https://doi.org/10.1016/j.biomaterials.2013.08.088>
- Hanks, S., Coleman, K., Reid, S., Plaja, A., Firth, H., FitzPatrick, D., Kidd, A., Méhes, K., Nash, R., and Robin, N., 2004. Constitutional aneuploidy and cancer predisposition caused by biallelic mutations in BUB1B. *Nat. Genet.*, **36**: 1159-1161. <https://doi.org/10.1038/ng1449>
- Heidel, J.D., Liu, J.Y.-C., Yen, Y., Zhou, B., Heale, B.S., Rossi, J.J., Bartlett, D.W., and Davis, M.E., 2007. Potent siRNA inhibitors of ribonucleotide reductase subunit RRM2 reduce cell proliferation *in vitro* and *in vivo*. *Clin. Cancer Res.*, **13**: 2207-2215. <https://doi.org/10.1158/1078-0432.CCR-06-2218>
- Hu, P., Chen, X., Sun, J., Bie, P., and Zhang, L., 2015. siRNA-mediated knockdown against NUF2 suppresses pancreatic cancer proliferation *in vitro* and *in vivo*. *Biosci. Rep.*, **35**. <https://doi.org/10.1042/BSR20140124>
- Huang, S., Qian, J., Yuan, B., Lin, Y., Ye, Z., and Huang, S., 2014. SiRNA-mediated knockdown against NUF2 suppresses tumor growth and induces cell apoptosis in human glioma cells. *Cell. mol. Biol.*, **60**: 30-36.
- Islami, F., Goding Sauer, A., Miller, K., Siegel, R., Fedewa, S., Jacobs, E., McCullough, M., Patel, A., Ma, J., Soerjomataram, I., Flanders, W., Brawley,

- O., Gapstur, S., and Jemal, A.J.C., 2018. Proportion and number of cancer cases and deaths attributable to potentially modifiable risk factors in the United States. *CA Cancer J. Clin.*, **68**: 31-54. <https://doi.org/10.3322/caac.21440>
- Jemal, A., Bray, F., Center, M., Ferlay, J., Ward, E., and Forman, D., 2011. Global cancer statistics. *CA Cancer J. Clin.*, **61**: 69-90. <https://doi.org/10.3322/caac.20107>
- Lee, E., Pain, M., Wang, H., Herman, J., Toledo, C., DeLuca, J., Yong, R., Paddison, P., and Zhu, J.J.C.R., 2017. BUB1B sensitivity to inhibition defines an alternative classification of glioblastoma. *Cancer Res.*, **77**: 5518-5529. <https://doi.org/10.1158/0008-5472.CAN-17-0736>
- Li, H., Gai, L., Wu, Z., and Li, F., 2022. Maternal embryonic leucine zipper kinase serves as a potential prognostic marker and leads to sorafenib chemoresistance modified by miR-142-5p in hepatocellular carcinoma. *Mol. Biol. Rep.*, **49**: 3015-3024. <https://doi.org/10.1007/s11033-022-07128-3>
- Liang, X., Dai, Y., Li, Z., Gan, M., Zhang, S., Yin-Pan, Lu, H., Cao, X., Zheng, B., Bao, L., Wang, D., Zhang, L., and Ma, S., 2014. Expression and function analysis of mitotic checkpoint genes identifies TTK as a potential therapeutic target for human hepatocellular carcinoma. *PLoS One*, **9**: e97739. <https://doi.org/10.1371/journal.pone.0097739>
- Liu, C., Yan, Y., Di, F., Li, W., Yin, X., and Dong, L., 2021. Inhibition of NCAPG expression inactivates the Wnt/ β -catenin signal to suppresses endometrial cancer cell growth *in vitro*. *Environ. Toxicol.*, **36**: 2512-2520. <https://doi.org/10.1002/tox.23364>
- Liu, C., Zhou, N., Li, J., Kong, J., Guan, X., and Wang, X.J.D.M., 2017a. Eg5 overexpression is predictive of poor prognosis in hepatocellular carcinoma patients. *Dis. Markers*, **2017**: 2176460. <https://doi.org/10.1155/2017/2176460>
- Liu, K., Li, Y., Yu, B., Wang, F., Mi, T., and Zhao, Y., 2018. Silencing non-SMC chromosome-associated polypeptide G inhibits proliferation and induces apoptosis in hepatocellular carcinoma cells. *Can. J. Physiol. Pharmacol.*, **96**: 1246-1254. <https://doi.org/10.1139/cjpp-2018-0195>
- Liu, W., Liang, B., Liu, H., Huang, Y., Yin, X., Zhou, F., Yu, X., Feng, Q., Li, E., Zou, Z., and Wu, L., 2017b. Overexpression of non-SMC condensin I complex subunit G serves as a promising prognostic marker and therapeutic target for hepatocellular carcinoma. *Int. J. mol. Med.*, **40**: 731-738. <https://doi.org/10.3892/ijmm.2017.3079>
- Liu, X., Liao, W., Yuan, Q., Ou, Y., and Huang, J., 2015. TTK activates Akt and promotes proliferation and migration of hepatocellular carcinoma cells. *Oncotarget*, **6**: 34309-34320. <https://doi.org/10.18632/oncotarget.5295>
- Malumbres, M., and Barbacid, M., 2005. Mammalian cyclin-dependent kinases. *Trends Biochem. Sci.*, **30**: 630-641. <https://doi.org/10.1016/j.tibs.2005.09.005>
- McGlynn, K.A., Petrick, J.L., and El-Serag, H.B., 2021. Epidemiology of hepatocellular carcinoma. *Hepatology*, **73**: 4-13. <https://doi.org/10.1002/hep.31288>
- Mills, G.B., Schmandt, R., McGill, M., Amendola, A., Hill, M., Jacobs, K., May, C., Rodricks, A.-M., Campbell, S., and Hogg, D., 1992. Expression of TTK, a novel human protein kinase, is associated with cell proliferation. *J. Biol. Chem.*, **267**: 16000-16006. [https://doi.org/10.1016/S0021-9258\(19\)49633-6](https://doi.org/10.1016/S0021-9258(19)49633-6)
- Mizukami, Y., Kono, K., Daigo, Y., Takano, A., Tsunoda, T., Kawaguchi, Y., Nakamura, Y., and Fujii, H., 2008. Detection of novel cancer-testis antigen-specific T-cell responses in TIL, regional lymph nodes, and PBL in patients with esophageal squamous cell carcinoma. *Cancer Sci.*, **99**: 1448-1454. <https://doi.org/10.1111/j.1349-7006.2008.00844.x>
- Nakano, I., Paucar, A., Bajpai, R., Dougherty, J., Zewail, A., Kelly, T., Kim, K., Ou, J., Groszer, M., Imura, T., Freije, W., Nelson, S., Sofroniew, M., Wu, H., Liu, X., Terskikh, A., Geschwind, D., and Kornblum, H., 2005. Maternal embryonic leucine zipper kinase (MELK) regulates multipotent neural progenitor proliferation. *J. Cell Biol.*, **170**: 413-427. <https://doi.org/10.1083/jcb.200412115>
- Nordlund, P., and Reichard, P.J.A., 2006. Ribonucleotide reductases. *Annu. Rev. Biochem.*, **75**: 681-706. <https://doi.org/10.1146/annurev.biochem.75.103004.142443>
- Park, H., Jeon, Y., Shin, H., Kim, I., Kang, H., Jeong, S., Chung, D., Lee, C.J.E., and Medicine, M., 2007. Differential promoter methylation may be a key molecular mechanism in regulating BubR1 expression in cancer cells. *Exp. mol. Med.*, **39**: 195-204. <https://doi.org/10.1038/emm.2007.22>
- Qian, X., Song, X., He, Y., Yang, Z., Sun, T., Wang, J., Zhu, G., Xing, W., and You, C., 2015. CCNB2 overexpression is a poor prognostic biomarker in Chinese NSCLC patients. *Biomed. Pharmacother.*, **74**: 222-227. <https://doi.org/10.1016/j.biomed.2015.03.005>

- biopha.2015.08.004
- Scintu, M., Vitale, R., Prencipe, M., Gallo, A., Bonghi, L., Valori, V., Maiello, E., Rinaldi, M., Signori, E., Rabitti, C., Carella, M., Dallapiccola, B., Altomare, V., Fazio, V., and Parrella, P.J.C.L., 2007. Genomic instability and increased expression of BUB1B and MAD2L1 genes in ductal breast carcinoma. *Cancer Lett.*, **254**: 298-307. <https://doi.org/10.1016/j.canlet.2007.03.021>
- Shannon, P., Markiel, A., Ozier, O., Baliga, N.S., Wang, J.T., Ramage, D., Amin, N., Schwikowski, B., and Ideker, T., 2003. Cytoscape: A software environment for integrated models of biomolecular interaction networks. *Genome Res.*, **13**: 2498-2504. <https://doi.org/10.1101/gr.1239303>
- Shao, J., Liu, X., Zhu, L., and Yen, Y.J.E., 2013. Targeting ribonucleotide reductase for cancer therapy. *Expert Opin. Therap. Targets*, **17**: 1423-1437. <https://doi.org/10.1517/14728222.2013.840293>
- Shichiri, M., Yoshinaga, K., Hisatomi, H., Sugihara, K., and Hirata, Y.J.C., 2002. Genetic and epigenetic inactivation of mitotic checkpoint genes hBUB1 and hBUBR1 and their relationship to survival. *Cancer Res.*, **62**: 13-17.
- Shin, H., Baek, K., Jeon, A., Park, M., Lee, S., Kang, C., Lee, H., Yoo, S., Chung, D., Sung, Y., McKeon, F., and Lee, C.J.C., 2003. Dual roles of human BubR1, a mitotic checkpoint kinase, in the monitoring of chromosomal instability. *Cancer Cell*, **4**: 483-497. [https://doi.org/10.1016/S1535-6108\(03\)00302-7](https://doi.org/10.1016/S1535-6108(03)00302-7)
- Singal, A.G., Lampertico, P., and Nahon, P., 2020. Epidemiology and surveillance for hepatocellular carcinoma: New trends. *J. Hepatol.*, **72**: 250-261. <https://doi.org/10.1016/j.jhep.2019.08.025>
- Sugimasa, H., Taniue, K., Kurimoto, A., Takeda, Y., Kawasaki, Y., and Akiyama, T., 2015. Heterogeneous nuclear ribonucleoprotein K upregulates the kinetochore complex component NUF2 and promotes the tumorigenicity of colon cancer cells. *Biochem. biophys. Res. Commun.*, **459**: 29-35. <https://doi.org/10.1016/j.bbrc.2015.02.043>
- Sung, H., Ferlay, J., Siegel, R. L., Laversanne, M., Soerjomataram, I., Jemal, A., and Bray, F., 2021. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.*, **71**: 209-249. <https://doi.org/10.3322/caac.21660>
- Tang, Z., Li, C., Kang, B., Gao, G., Li, C., and Zhang, Z., 2017. GEPIA: A web server for cancer and normal gene expression profiling and interactive analyses. *Nucl. Acids Res.*, **45**: W98-W102. <https://doi.org/10.1093/nar/gkx247>
- Wan, X., Yeung, C., Kim, S., Dolan, J., Ngo, V., Burkett, S., Khan, J., Staudt, L., and Helman, L.J.C., 2012. Identification of FoxM1/Bub1b signaling pathway as a required component for growth and survival of rhabdomyosarcoma. *Cancer Res.*, **72**: 5889-5899. <https://doi.org/10.1158/0008-5472.CAN-12-1991>
- Wang, L., Huang, J., Jiang, M.J.C., 2010. RRM2 computational phosphoprotein network construction and analysis between no-tumor hepatitis/cirrhotic liver tissues and human hepatocellular carcinoma (HCC). *Biochem. Int. J. Exp. Cell. Physiol. Pharmacol.*, **26**: 303-310. <https://doi.org/10.1159/000320553>
- Wang, Y., Gao, B., Tan, P., Handoko, Y., Sekar, K., Deivasigamani, A., Seshachalam, V., OuYang, H., Shi, M., Xie, C., Goh, B., Ooi, L., and Man Hui, K., 2019a. Genome-wide CRISPR knockout screens identify NCAPG as an essential oncogene for hepatocellular carcinoma tumor growth. *FASEB J. Off. Publ. Fed. Am. Soc. Exp. Biol.*, **33**: 8759-8770. <https://doi.org/10.1096/fj.201802213RR>
- Wang, Y., Tan, P., Handoko, Y., Sekar, K., Shi, M., Xie, C., Jiang, X., Dong, Q., Goh, B., Ooi, L., Gao, Z., and Hui, K., 2019b. NUF2 is a valuable prognostic biomarker to predict early recurrence of hepatocellular carcinoma after surgical resection. *Int. J. Cancer*, **145**: 662-670. <https://doi.org/10.1002/ijc.32134>
- Warde-Farley, D., Donaldson, S.L., Comes, O., Zuberi, K., Badrawi, R., Chao, P., Franz, M., Grouios, C., Kazi, F., and Lopes, C.T., 2010. The GeneMANIA prediction server: Biological network integration for gene prioritization and predicting gene function. *Nucl. Acids Res.*, **38**: W214-W220. <https://doi.org/10.1093/nar/gkq537>
- Wu, B., Hu, C., Kong, L.J.E., and Medicine, T., 2021. ASPM combined with KIF11 promotes the malignant progression of hepatocellular carcinoma via the Wnt/ β -catenin signaling pathway. *Exp. Therap. Med.*, **22**: 1154. <https://doi.org/10.3892/etm.2021.10588>
- Xia, H., Kong, S., Chen, J., Shi, M., Sekar, K., Seshachalam, V., Rajasekaran, M., Goh, B., Ooi, L., and Hui, K., 2016. MELK is an oncogenic kinase essential for early hepatocellular carcinoma recurrence. *Cancer Lett.*, **383**: 85-93. <https://doi.org/10.1016/j.canlet.2016.09.017>
- Xie, Y., Wang, A., Lin, J., Wu, L., Zhang, H., Yang, X., Wan, X., Miao, R., Sang, X., and Zhao, H., 2017. Mps1/TTK: A novel target and biomarker for cancer. *J. Drug Target.*, **25**: 112-118. <https://doi.org/10.1080/1061186X.2016.1258568>

- Xu, Q., Ge, Q., Zhou, Y., Yang, B., Yang, Q., Jiang, S., Jiang, R., Ai, Z., Zhang, Z., and Teng, Y., 2020. MELK promotes Endometrial carcinoma progression via activating mTOR signaling pathway. *EBio Med.*, **51**: 102609. <https://doi.org/10.1016/j.ebiom.2019.102609>
- Yam, C.H., Fung, T.K., and Poon, R.Y., 2002. Cyclin A in cell cycle control and cancer. *Cell Mol. Life Sci.*, **59**: 1317-1326. <https://doi.org/10.1007/s00018-002-8510-y>
- Yang, J.D., and Heimbach, J.K., 2020. New advances in the diagnosis and management of hepatocellular carcinoma. *Br. Med. J.*, **371**. <https://doi.org/10.1136/bmj.m3544>
- Yang, J.D., Hainaut, P., Gores, G.J., Amadou, A., Plymoth, A., and Roberts, L.R., 2019. A global view of hepatocellular carcinoma: Trends, risk, prevention and management. *Nat. Rev. Gastroenterol. Hepatol.*, **16**: 589-604. <https://doi.org/10.1038/s41575-019-0186-y>
- Yang, N., Eijssink, J.J., Lendvai, Á., Volders, H.H., Klip, H., Buikema, H.J., van Hemel, B.M., Schuurings, E., van der Zee, A.G., and Wisman, G.B.A., 2009. Methylation markers for CCNA1 and C13ORF18 are strongly associated with high-grade cervical intraepithelial neoplasia and cervical cancer in cervical scrapings. *Cancer Epidemiol. Prev. Biomark.*, **18**: 3000-3007. <https://doi.org/10.1158/1055-9965.EPI-09-0405>
- Yang, P., Lin, L., and Liu, T.J.B., 2020. Sorafenib inhibits ribonucleotide reductase regulatory subunit M2 (RRM2) in hepatocellular carcinoma cells. *Biomolecules*, **10**. <https://doi.org/10.3390/biom10010117>
- Zhang, P., Feng, J., Wu, X., Chu, W., Zhang, Y., and Li, P.J.P., 2021. Bioinformatics analysis of candidate genes and pathways related to hepatocellular carcinoma in China: A study based on public databases. *Pathol. Oncol. Res.*, **27**: 588532. <https://doi.org/10.3389/pore.2021.588532>
- Zhang, Q., Su, R., Shan, C., Gao, C., and Wu, P., 2018. Non-SMC condensin I complex, subunit G (NCAPG) is a novel mitotic gene required for hepatocellular cancer cell proliferation and migration. *Oncol. Res.*, **26**: 269. <https://doi.org/10.3727/096504017X15075967560980>
- Zhang, X., Wang, H., Han, Y., Zhu, M., Song, Z., Zhan, D., and Jia, J., 2020a. NCAPG induces cell proliferation in cardia adenocarcinoma via PI3K/AKT signaling pathway. *Oncol. Targets Ther.*, **13**: 11315-11326. <https://doi.org/10.2147/OTT.S276868>
- Zhang, X., Wang, J., Wang, Y., Liu, G., Li, H., Yu, J., Wu, R., Liang, J., Yu, R., and Liu, X., 2020b. MELK inhibition effectively suppresses growth of glioblastoma and cancer stem-like cells by blocking AKT and FOXM1 pathways. *Front. Oncol.*, **10**: 608082. <https://doi.org/10.3389/fonc.2020.608082>
- Zhou, B., Tsai, P., Ker, R., Tsai, J., Ho, R., Yu, J., Shih, J., Yen, Y.J.C., and Metastasis, E., 1998. Overexpression of transfected human ribonucleotide reductase M2 subunit in human cancer cells enhances their invasive potential. *Clin. exp. Metastasis*, **16**: 43-49. <https://doi.org/10.1023/A:1006559901771>
- Zhou, Y., Zhou, B., Pache, L., Chang, M., Khodabakhshi, A.H., Tanaseichuk, O., Benner, C., and Chanda, S.K., 2019a. Metascape provides a biologist-oriented resource for the analysis of systems-level datasets. *Nat. Commun.*, **10**: 1-10. <https://doi.org/10.1038/s41467-019-09234-6>
- Zhou, Z., Li, Y., Hao, H., Wang, Y., Zhou, Z., Wang, Z., and Chu, X.J.C., 2019b. Screening hub genes as prognostic biomarkers of hepatocellular carcinoma by bioinformatics analysis. *Cell Transpl.*, **28**: 76S-86S. <https://doi.org/10.1177/0963689719893950>